ATTACHMENT 6 TITLE OF THE INVENTION

DECREASED FAT ABSORPTION WITH AN ANTI-LIPASE ANTIBODY

BACKGROUND OF THE INVENTION

Field of the invention

A food additive that decrease fat absorption in mammals

Discussion of the background

Our sedentary life including the decreased physical activity and increased food intake have made us prone to be overweight. The above has brought as consequence that almost 40-50 % of the USA population is 20% above their desirable weight. The advance in the science of food and nutrition not only has made us wiser about the functions of all nutrients but also by applying that knowledge we have concentrated food in smaller portions by which the satisfaction of filling is decreased. Even if the amount of food intake remains the same, we will have an excess caloric intake due to the high energy concentration of such type of food (Bell, et al 1997). Currently, the weight loss related market is full of diet

pills that reduce appetite by suppressing brain hormones, drugs that suppress the absorption of nutrients, pills that supposedly have ergogenics effect, pills that increase food passage rate and other fad diets. Mostly all of these drugs have secondary effects like depression, anxiety, addiction and others.

A new approach for the reduction of calories in food is by the use of fat substitutes (Gershoff, et al 1995). Each gram of fat provides 9 calories as compared to 4 calories per gram of carbohydrate and protein. Fat substitutes mainly those made of long carbohydrate chains are use for the elaboration of prepared food with the purpose of maintaining fat properties in the prepared food but decreasing calories. A new fat substitute, Olestra, which is made of long chain fatty acids that are too big for digestive enzymes (lipase) to breakdown, therefore that type of fat is not absorbed. It has been observed that the consumption of Olestra has resulted in decreased absorption of fat soluble and the presence of fat in the feces. A long term study (12 weeks) where 1/3 of the dietary fat was replaced with olestra, female subjects lost weight and did not compensate for the reduced calories and fat intake (Roy, et al, 1997).

In the animal industry, researchers have been working on the reduction of fat accumulation in animals since this characteristic first, has a negative effect on profits and second, consumers want less visible fat in order to decrease the health risk.

Fat accumulation in animals has been reduced by passively administered antibodies against adipocyte plasma membrane in rats, pigs, rabbits and lambs. Immunity against growth hormone has also decreased abdominal fat in chickens (Brodie and Hu, 1996; Moloney, 1995; Flint, 1992).

Lipase, an enzyme produced by the pancreas, hydrolyzes triacylglicerides into free fatty acids and glycerol. This is a crucial step in breaking down ingested fat in the gastro-intestinal tract. Lipase is more active in the duodenum (small intestine) where broken down fat with the aid of bile salts form micelles and then are absorbed by the intestinal mucosa.

Therefore, by inhibiting lipase the ingested fat will not be absorbed and the energy supplied by fat and the fat itself will be excreted.

SUMMARY OF THE INVENTION

A method for the inhibition of fat absorption in mammals.

DESCRIPTION OF THE PREFERRED EMBODIMENTS

The present invention relates to a method for decreasing fat absorption by orally feeding chicken antibodies against lipase. The preferred antigen for obtaining the antibodies is a swine pancreatic extract that contains lipase. This antigen is commercially produced by Sigma Chemical Co. Lipase is a conserve molecule with similar structure between animal and plant species, therefore an antibody against swine lipase will cross-react with other species' lipase. We have found that by feeding anti-lipase antibodies to mice and rats will result in either decreased body weight or reduced feed efficiency. The antibody extract can either be fed in water suspension, included in feed as dry powder and/or encapsulated in liposomes.

Previous research on the effectiveness of chicken antibodies has been reported; i.e. the prevention of bacterial infection in swine, calf and dairy cows (Yokoyama et al, 1993; Erhard et al 1993; Coleman, 1995). These

researches have also demonstrated the presence of intact avian antibodies in the gastro-intestinal tract of the animals.

Although chickens antibodies are known to protect against bacterial infections, no antibody has been reported to decreased fat absorption.

It will be apparent for those skilled in the art that the aforementioned objects and other advantages may be further achieved by the practice of the present invention.

Example 1

This example illustrate the preparation of the specific antibody against lipase. 17-week old hens were injected with 2.5 mg of lipase (Sigma Chemical Co.). The inoculum was prepared by dissolving the enzyme in 0.2 ml phosphate buffered saline (PBS, pH 7.3) and 0.2 ml complete Freund's adjuvant. The antigen preparation was injected into two sites 0.2 ml in each (right and left) pectoralis muscle. A total of 0.4 ml of antigen preparation per hen was administered. A second injection was administered 5-6 weeks following the initial injection (at about 50% egg hen production). In the second antigen preparation, incomplete Freund's adjuvant was used instead of complete Freund's adjuvant. Hens were re-

injected with the antigen preparation every two months or when the antibody titer was determined to be low. Antibody titer was determined by ELISA. Hens had free access to fed and water and they were maintained in an isolated room in order to minimized outside contamination.

Example 2

Antibody was purified as follows: One volume of egg yolk of example 1 was mixed with 9 volumes of distilled water and left to sit overnight at 4 °C. Then the aqueous portion was centrifuged at 4000 rpm for 10 minutes and filtered through a cheesecloth in order to remove any excess fat. The aqueous portion contains all the protein present in the egg yolk which includes all the antibodies (IgY). The liquid was frozen and then was freeze dried. The antibody activity was determine by ELISA.

Example 3

Antibody against lipase was determined as follows:

1.- ELISA plates were coated with 100 ul lipase preparation (50 ug/ml) in carbonate buffer. The plates were incubated at 4 °C overnight prior to blocking with 1.5% bovine serum albumin for 4 hours at room temperature.

2.- 100 ul of a 0.5 mg protein/ml antibody extract was added to each well and the plates incubated at room temperature for 1 hour.

3.- Plates were washed with PBS-tween solution. 100 ul of rabbit anti-chicken IgG conjugated to horseradish peroxidase was added to each well. The plates were incubated at room temperature for 1 hour.

4.- Plates were washed with PBS-tween and 100 ul of TMB substrate was added to each well and incubated for 15 minutes.

5.- The reaction was stopped with 100 ul of 2 M sulfuric acid.

6.- Plates were read at 455 nm in an ELISA plate reader.

7.- Titer was determine as the inverse of the dilution at which O.D. of the immunized egg was similar to the un-immunized control (O.D. < 0.100).

Example 4

This study illustrates the in vitro inhibition of lipase by the chicken anti-lipase antibody. The effectiveness of the antibody was verified by using a test specific for the determination of lipase in serum (Sigma Chemical Co). We modified this test by adding a known amount of enzyme

(lipase) and antibody to a certain volume of phosphate buffered saline. The resulting activity was expressed as Sigma-Tietz units/ml, which is equal to the ml of 0.05 N NaOH required to neutralize the fatty acid formed in the reaction. In a preliminary study we found the following:

Lipase (mg)	**anti-lipase (protein extract) (mg)**	**Lipase Activity (U)**	**% decreased activity**
2.0	0	17.3	
2.0	9.0	18.8	0
1.0	0	14.0	
1.0	9.0	12.5	11
0.5	0	10.4	
0.5	9.0	9.8	6
0.25	0	8.1	
0.25	9.0	6.7	17

In a second test; higher amount of antibody extract was used. The results are as follows:

Lipase (mg)	Anti-lipase Protein Extract (mg)	Lipase Activity (U)	% decreased activity
2.0	0	18.7	
2.0	37	14.0	25
1.0	0	13.5	
1.0	37	6.9	49

Example 5

This study illustrates the effect of anti-lipase antibody in mice. Two groups of 5 2-month old mice (25-34 gr each) were given 5 mg of antibody (protein extract) per ml of water. The antibody was mixed with water on a daily basis. Mice were fed the same amount of feed in both groups (approx. 5 gr/mice/day). The length of the experiment was 58 days. The results are as follows

	total* initial body weight (gr)	total final body weight (gr)	difference in body weight (gr)	total feed intake (gr)	gr of feed needed to gain 1 gr of body weight
control	157	199	42	1039	24.74
anti-lipase	156	187	31	1039	33.52

* Sum of 5 mice/trt

Example 6

This study illustrate the effect of anti-lipase when fed at lower levels than in example 5. Two groups of 5 5-month old mice (32-35 gr each) were given 1 mg of antibody (protein extract) per ml of water for the first 7 days then it was increased to 2 mg/ml of water. Mice in both groups were fed the same amount of feed for 35 days. The results were as follows.

	Total initial body weight (gr)	**Total final body weight (gr)**	**difference in body weight**	**total feed intake (gr)**	**gr of feed needed to gain 1 gr of body weight**
control	182	204	22	787.4	35.79
anti-lipase	181	185	4	788.5	197.13

Example 7

This study demonstrate the encapsulation of anti-lipase antibodies by liposomes. This liposome preparation was based on the procedure by Shimizu et al (1993). Final liposome suspension was frozen and later freeze dried. A known amount

of freeze dried liposome was mixed with rat diet, and fed daily for the length of the study.

Example 8

This study illustrate the effect of anti-lipase antibodies in rats. Twelve retired breeder Sprague Dawley rats (Harlan, Wisconsin) were individually caged and supplied with free access to water. They were fed a rabbit chow which was supplemented with corn oil in order increase fat content to 30%. Feed intake was monitored for 1 week in order to determine the amount of feed needed to maintain their initial body weight. Rats were divided in two groups one fed the high fat diet and the other group was fed the same diet with freeze dried liposome containing anti-lipase antibody extract. The treated diet contained 750 mg antibody/kg of diet. The results after 1 week of treatment are as follows:

	Initial body weight (gr)*	**One week feed intake (gr)**	**Final body weight (gr)**	**grams of feed needed to gain 1 gr of body weight**
control	316	132.3	327	12.0
antibody	319	129.4	326	18.5

* average of 6 rats.

Example 9

Since it was observed that rats gained weight in example 8, the same rats were used but this time feed was restricted. The results are as follows:

	initial body weight (gr)	**Final body weight (gr)**	**difference in body weight**	**feed intake (gr)**
control	327	319	-8	102
antibody	326	317	-9	101

Example 10

This study demonstrate the effect of the anti-lipase when fed to rats at maintenance feed intake. The results are as follows:

	initial body weight (gr)	**One week body weight (gr)**	**difference in body weight**	**One week feed intake (gr)**	**gr of feed needed to gain 1 gr of body weight**
control	325	332	7	112	16
antibody	324	325	1	114	114

It will be apparent to those skilled in the art that a number of modifications and variations may be made without departing from the scope of the present invention as set forth in the appended claims.

REFERENCES

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What is claimed is :

1.- A method for decreasing fat absorption in mammals and avian by feeding an antibody that binds lipase.

2.- A method of claim 1 wherein lipase is any enzyme that is needed for the hydrolysis of fat in order for it to be absorbed by the gastro-intestinal mucosa.

3.- A method of claim 1 wherein the antibody binds to lipase therefore inhibiting its activity in the gastro intestinal tract.

4.- A method of claim 1 wherein lipase is of mammal, avian or plant origin.

5.- A method of claim 4 wherein mammal is such as human, primates, monogastrics and ruminants.

6.- A method of claim 4 wherein avian is such as chicken, turkey, goose, duck, quail, pheasant, pigeon.

7.- A method of claim 4 wherein plants include bacteria, mold and yeast.

8.- A method of claim 1 wherein antibody was produced in avian eggs.

9.- A method of claim 1 wherein the antibody can also be produced in other commercially or laboratory antibody-

producing animal including monoclonal, plant and bacteria produced antibodies.

10.- A method of claim 8 wherein avian compromise chicken, duck, goose, turkey, pheasant, quail, pigeon.

11.- A method of claim 8 wherein the antibodies are obtained from unfractionated whole eggs.

12.- A method of claim 8 wherein the antibodies are obtained from the yolk of an egg without fractionation thereof.

13.- A method of claim 8 wherein the antibodies are obtained by fractionating the egg yolk resulting in a protein concentrate or pure IgY (chicken immunoglobulin).

14.- A method of claim 1 wherein antibody produced as claim 8-13 is kept as it was obtained or is further processed in order to freeze dry, spray dry or encapsulated.

15.- A method of claim 14 wherein encapsulation is such process that protect the antibody against changes that inactivate or disrupt the effectiveness of the antibody.

16.- A method of claim 14 wherein encapsulation methods are liposomes, protein coating, carbohydrate coating, other chemical processes that will coat the antibody.

17.- A method of claim 1 wherein the antibody or the antibody containing material is orally fed.

18.- A method of claim 17 wherein the orally fed antibody or the antibody containing material is fed by itself as powder form, liquid form, compressed tablet or other type of pill/tablet like material.

19.- A method of claim 18 wherein the powder or the liquid antibody or the antibody containing material are fed as part of a processed or prepared food.

20.- A method of claim 19 wherein processed or prepared food is any food were the antibody or the antibody containing material in powder or liquid form have been included as part of the formulation or recipe.

21.- A method of claim 20 wherein processed or prepared food is any food for human or animal consumption which include ready to eat, ready to mix, concentrate, additives, refrigerated and frozen food.

22.- A method of claim 1 wherein decreased fat absorption is due to the decreased lipase activity, therefore fat is excreted and not absorbed in the gastro-intestinal tract.

23.- A method of transferring gastro-intestinal enzymes antibodies in animals and humans to other animals or humans in order to decrease absorption of nutrients such as protein, carbohydrates and lipids.

24.- A method of claim 23 wherein said gastro-intestinal tract enzymes is selected from amylase, trypsin, chymotrypsin, protease and other enzymes required for the absorption of nutrients.

25.- A method of transferring antibodies from an animal or human, to other animals or humans in order to modify a biochemical process comprising:

administering to said animal or human an antibody containing substance wherein said substance is derived from a producer animal or human wherein said producer animal or human has been immunized with the antigen wherein said antigen regulate a biochemical process in the gastro-intestinal tract.

ABSTRACT OF THE DISCLOSURE

A method for the decrease of fat absorption in any animal, wherein the animal is fed an antibody produced against lipase, an enzyme which is required for fat absorption.